

长花龙血树茎中抗氧化活性的木脂素类成分研究

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摘要 从长花龙血树的茎中分离得到一个新的木脂素化合物, 命名为 angudracanolide A (**1**), 以及三个已知的木脂素 **2~4** 和三个已知的苯基丙酸类化合物 **5~7**. 通过对光谱数据的分析和量子化学计算, 阐明了新化合物的绝对构型. 与作为阳性对照的 trolox 相比(IC₅₀ 值为 0.45 mmol/L), 化合物 **1~4** 显示出明显的抗氧化活性, IC₅₀ 值分别为 15, 19, 25, 30 mmol/L.

关键词 长花龙血树; 木脂素; 抗氧化活性

Lignanols with Antioxidant Activity from the Stems of *Dracaena angustifolia*

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Abstract A new lignanolide given a trivial name angudracanolide A (**1**), together with three known lignans **2~4** and three known phenylpropanoids **5~7**, was isolated from the stems of *Dracaena angustifolia*. The complete structure of new compound was elucidated by analysis of spectroscopic data and quantum chemistry calculations. Compounds **1~4** showed obvious antioxidant activity with IC₅₀ values of 15, 19, 25, 30 mmol/L, respectively by comparing the standard trolox with a IC₅₀ value of 45 mmol/L.

Keywords *Dracaena angustifolia*; lignan; antioxidant activity

1 Introduction

Dracaena angustifolia was composed of more than 40 species and mainly found in Hainan, Taiwan and Yunnan

provinces, in China, as well as Australia, India, Malaysia, and Philippines.^[1] *D. angustifolia* is also regarded as one of the Li-nationality medicinal plants for treatment of endemic arthralgia and myalgia on Hainan Island, China. Hitherto

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there are few research reports about the chemical constituents of *D. angustifolia*. Our previous phytochemical studies on the species of *D. angustifolia* had led to the discovery of a type of phenolic compounds.^[2] Recently, investigation of the ethyl acetate extract from *D. angustifolia* afforded seven compounds, including a new lignanolide angudracanolide A (**1**), and three known lignans (–)-syringaresinol (**2**),^[3] de-4'-*O*-methyl-yangambin (**3**),^[4] (+)-isolariciresinol (**4**),^[5] along with three known phenylpropanoids zhebeiresinol (**5**),^[6] β -hydroxypropio-vanillone (**6**),^[7] erythro-7*R*,8*S*-7-*O*-ethylguaiacyl glycerol (**7**)^[8] (Figure 1). Meanwhile, the antioxidant activity of all compounds above was also evaluated.

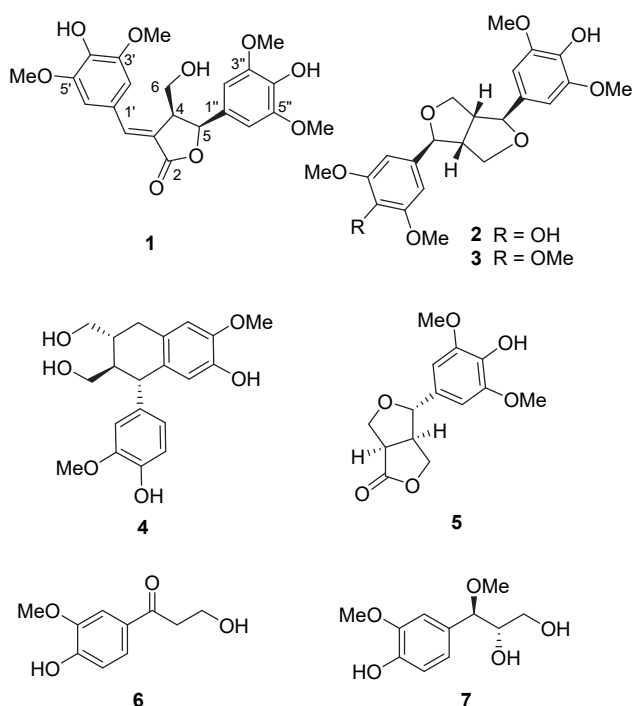


Figure 1 Chemical constituents from *Dracaena angustifolia*

2 Results and discussion

2.1 Structure elucidation

Compound **1** was obtained as a white amorphous powder with a molecular formula of $C_{22}H_{24}O_9$ by high resolution electrospray ionization mass spectroscopy (HRESIMS) spectrum showing $[M+H]^+$ ion at m/z 433.1495 (calcd for 433.1493), suggesting eleven degrees of unsaturation.

The 1H NMR spectrum of **1** showed an olefinic proton signal at δ_H 7.49 (s, 1H, H-7'), two groups of equivalent protons at δ_H 6.99 (s, 2H, H-2',6') and 6.55 (s, 2H, H-2'',6''), two pairs of equivalent methoxy groups at δ_H 3.80 (s, 6H, H-3'',5''-OMe) and 3.73 (s, 6H, H-3',5'-OMe), two sp^3 methine protons at δ_H 5.55 (s, 1H, H-5) and 3.63~3.66 (m, 1H, H-4), and an oxygenated methylene signal with chemical shifts of δ_H 3.85~3.89 (m, 1H, H-6) and 3.60~3.64 (m, 1H, H-6). The ^{13}C NMR spectrum (Table 1) presented 22 carbon signals containing an ester carbonyl carbon at δ_C

Table 1 1H NMR (400 MHz) and ^{13}C NMR (100 MHz) data of compound **1** in DMSO- d_6

Position	1	
	δ_H (J in Hz)	δ_C
2		171.9
3		131.0
4	3.63~3.66, m	49.6
5	5.55, s	81.0
6	3.60~3.64, m; 3.85~3.89, m	61.1
1'		123.8
2',6'	6.99, s	108.4
3',5'		148.1 ^a
4'		138.4
7'	7.49, s	138.5
1''		122.2
2'',6''	6.55, s	103.2
3'',5''		148.2 ^a
4''		135.7
3',5'-OMe	3.73, s	56.1
3'',5''-OMe	3.80, s	56.2

^a Interchanged.

171.9, nine sp^2 none-protonated carbons including two groups of equivalent carbons at δ_C 148.2, 148.1, five sp^2 methine carbons comprised of two suites of isovalent carbons at δ_C 108.4 and 103.2, and a carbon at δ_C 138.5, as well as two batteries of equivalent methoxyl carbons at δ_C 56.2 and 56.1, three sp^3 carbons consisted of an oxygenated methylene at δ_C 61.1, and two methines with chemical shifts of δ_C 81.0 and 49.6. Analysis of the 1H NMR and ^{13}C NMR data of **1**, revealed that it was extremely similar to those of descurnolide B,^[9] except for the additional appearances of a methylol group (δ_C 61.1) and a methine (δ_C 49.6) in **1**, and the absence of a methylene (δ_C 35.4) in descurnolide B.^[9] Thus, it provided a speculation that the structure of **1** was an analog of descurnolide B^[9] with a methylol located at C-4 (δ_C 49.6). This was supported by the observation of HMBC correlations. In the HMBC spectrum, the observed HMBC correlations from H-5 (δ_H 5.55) to C-2 (δ_C 171.9)/C-3 (δ_C 131.0)/C-4 (δ_C 49.6)/C-6 (δ_C 61.1), from H-7' (δ_H 7.49) to C-2 (δ_C 171.9)/C-4 (δ_C 49.6), from H-4 (δ_H 3.66) to C-3 (δ_C 131.0)/C-5 (δ_C 81.0)/C-6 (δ_C 61.1), ascertained that the methylene CH₂-4 in descurnolide B^[9] was substituted by the methylol group (δ_H 3.87/3.63, δ_C 61.1) in **1**. The geometrical configuration of the double bond between C-3 (δ_C 131.0) and C-7' (δ_C 138.5) was determined as *E* by NOESY experiment, which showed a cross-peak between H-4 and H-2' (Figure 2). However, the establishment of the relative configuration of **1** was challenged by observation of NOESY spectrum, due to H-4 was in close proximity to H-5 in the furan ring. Thus, the relative configuration of **1** was determined by DP4+ probability analysis.^[10] Structure of

1 with two chiral centers afforded two possible candidate diastereoisomers **1a** ($4R^*,5S^*$ -**1**) and **1b** ($4S^*,5S^*$ -**1**).

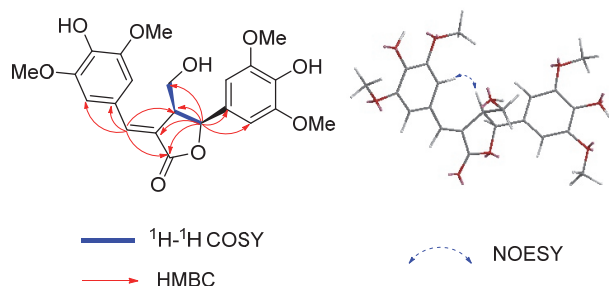


Figure 2 Key HMBC ^1H - ^1H COSY and NOESY correlations for compound **1**

Subsequently, diastereoisomers **1a** and **1b** were employed for systematic conformational analysis by Spartan' 14 software with molecular mechanic force field (MMFF94) method, respectively. The yielded conformations for **1a** and **1b** were performed structural optimizations and single-point energy calculations. The conformations with Boltzmann Gibbs energy distributions above 1% were used for calculations of NMR data, respectively. As a result, diastereoisomer **1b** with $4S^*/5S^*$ configuration was in high confidence with a DP4+ probability value of 100% relative to **1a**. Therefore, the relative configuration of **1** was assigned as $4S^*,5S^*$ -**1**. Further the absolute configuration of **1** was established by comparison of its calculated electrostatic circular dichroism (ECD) spectra with the experimental one. Finally, a comprehensive ECD spectrum for $4S,5S$ -**1** was calculated on the PBE1PBE/6-311g(d) (SMD) level of theory, while a mirrored ECD spectrum for the enantiomer $4R,5R$ -**1** was also afforded. And the absolute configuration of **1** was regarded as $4S,5S$ because the calculated curve of $4S,5S$ -**1** matched the experimental spectrum well (Figure 3).

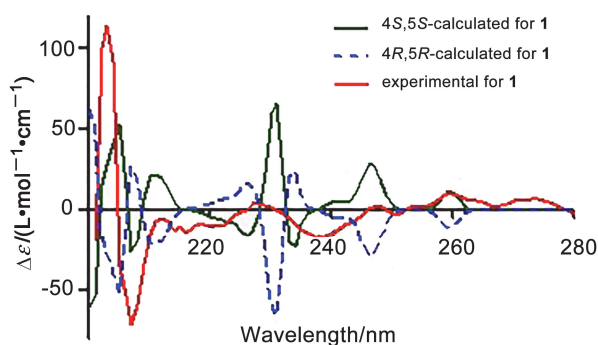


Figure 3 Comparison of the calculated and experimental ECD spectra for compound **1**

Based on above, the complete structure of **1** was established as $4S,5S,3E$ -(4-hydroxy-3,5-dimethoxy-benzylidene)-4-methylol-5-(4-hydroxy-3,5-dimethoxy-phenyl)-2(2*H*)-furanone, and was given a trivial name angudracanolide A, as depicted in Figure 1.

Other six known (–)-syringaresinol (**2**),^[3] de-4'-*O*-

methylyangambin (**3**),^[4] (+)-isolariciresinol (**4**),^[5] zhebeiresinol (**5**),^[6] β-hydroxypropiovanillone (**6**),^[7] erythro-7*R*,8*S*-7-*O*-ethylguaiaacylglycerol (**7**)^[8] (Figure 1) were identified by comparing their physical and spectroscopic data with those of compounds in literatures.

2.2 Antioxidant activity

Compounds **1**~**7** were evaluated for antioxidant activity, while trolox was used as a positive control. Among them, compounds **1**~**4** showed obvious antioxidant activity with IC_{50} values of 0.15, 0.19, 0.25 and 0.30 mmol/L, respectively, relative to positive control trolox with a IC_{50} value of 0.45 mmol/L (Table 2).

Table 2 Antioxidant activity of compounds **1**~**7**

Compd.	$\text{IC}_{50}/(\text{mmol}\cdot\text{L}^{-1})$
1	15
2	19
3	25
4	30
5	>50
6	>50
7	>50
Trolox	45

3 Conclusions

In this study, a new lignanolide named angudracanolide A (**1**), and three known lignans **2**~**4**, along with three known phenylpropanoids **5**~**7** were isolated from the stems of *D. angustifolia*. It was noted that compounds **4**~**6** were got from the species of *D. angustifolia* for the first time. In addition, compounds **1**~**4** showed obvious antioxidant activity with respect to the positive control trolox. This investigation enriches the structural diversity of *D. angustifolia* and provides good molecular templates for the development of new drug lead compounds.

4 Experimental section

4.1 General experimental procedures

IR spectra were recorded on a Nicolet 6700 spectrophotometer. Optical rotations were measured on a JASCO P-1020 digital polarimeter. ECD spectra were obtained at room temperature on a Jasco J-815 circular dichroism spectrometer. NMR spectra were recorded on a Bruker AV spectrometer (400 MHz for ^1H NMR and 100 MHz for ^{13}C NMR). Tetramethylsilane (TMS) was used as a reference. HRESIMS spectra were measured on a Q-TOF Ultima Global GAA076 LC mass spectrometer. Semi-preparative high performance liquid chromatography (HPLC) was performed on an Agilent 1260 LC series with a DAD detector using an Agilent Eclipse XDB-C₁₈ column (9.4 mm×250 mm, 5 μm). Silica gels (Qing Dao Hai Yang Chemical Group Co.; 100~200, 200~300 mesh) were

used for open column chromatography (CC). Pre-coated silica gel plates (Yan Tai Zi Fu Chemical Group Co.; G60, F-254) were used for thin-layer chromatography (TLC). Microplate spectrophotometer (BD, Bioscience U. S. A.) was used as a template reader for testing antioxidant activity.

4.2 Plant material

Dracaena angustifolia was collected in July 2017 from Changjiang city, Hainan province, China. The species was authenticated as *Dracaena angustifolia* by Dr. Yu-Kai Chen (Hainan Normal University, Hainan, China). The specimens (No C17-L07) were deposited at the Key Laboratory of Tropical Medicinal Resource Chemistry of Ministry of Education, Hainan Normal University (Hainan, China).

4.3 Extraction and isolation

The powdered air-dried stems (10 kg) of *D. angustifolia* were extracted with 75% ethanol (20 L×4, 5 d each) at room temperature. The filtered solution was evaporated in vacuum to give a crude extract. Subsequently, the extract was partitioned with petroleum ether and ethyl acetate solvent, respectively. The extract (90 g) derived from ethyl acetate layer was separated using a normal phase silica gel column chromatography (CC) [petroleum ether/ethyl acetate and chloroform/methanol, step gradient], affording ten major fractions (Fr.1~Fr.10). Frs.8 (9.9 g) was subjected to silica gel column chromatography (CC) (petroleum ether/ethyl acetate and chloroform/methanol, step gradient), attaining nine sub-fractions (Frs.8.1~Frs.8.9). Frs.8.2 (1.1 g) was further purified using semi-preparative HPLC (MeCN/H₂O, *V*:*V*=30:70) to obtain **5** (2.4 mg) and **6** (9.7 mg). Frs.8.6 (2.6 g) was prepared through semi-preparative HPLC (MeCN/H₂O, *V*:*V*=15:85, ULTRON ES-OVM column) to obtain **1** (13.4 mg) and **4** (17.4 mg). Frs.8.7 (5.0 g) was subjected to CC (chloroform/methanol from 30% to 100%) to yield three sub-fractions (Fr.8.7.1~Fr.8.7.3). Fr.8.7.1 (2.1 g) was purified by CC (petroleum ether/acetone, *V*:*V*=7:3~1:1) to afford compound **7** (3.81 mg). Frs.8.7.2 (2.5 g) was performed by reverse-phase (RP) CC and eluted with methanol/water (from 10% to 100%), obtaining three subfractions (Fr.8.7.2.1~Fr.8.7.2.3). Fr.8.7.2.2 (200 mg) was separated by repeated semi-preparative HPLC (MeOH/H₂O, *V*:*V*=30:70) to obtain **2** (5.1 mg). Fr.8.7.2.3 (1.4 g) was purified by semi-preparative HPLC (MeCN/H₂O, *V*:*V*=10:90, ULTRON ES-OVM column) to collect **3** (12.4 mg).

4.4 Physical properties of compound 1

Compound **1**: White amorphous powder; [α]_D²⁵ −3.5 (*c* 0.1 MeOH); UV (MeOH) $\lambda_{\max}$ [log ϵ /(L·mol^{−1}·cm^{−1})]: 338.0 (0.810), 242.0 (0.837), 208.0 (2.21) nm; IR (KBr) $\nu_{\max}$: 3554, 3437, 2939, 2846, 1734, 1629, 1605, 1516, 1461, 1334, 1213, 1157, 1114, 1043, 831, 735, 681, 624, 565, 527 cm^{−1}; ECD (*c* 1.16 mmol/L, MeOH) $\lambda_{\max}$ [$\Delta\epsilon$ /(L·mol^{−1}·cm^{−1})]: 203 (114.06), 212 (−5.11), 227 (3.27) nm. HR-ESI-MS calcd for C₂₂H₂₅O₉ [M+H]⁺ 433.1495, found 433.14934.

4.5 Assay of antioxidant activity

The antioxidant capacity was measured using T-AOC assay kit.^[11-12] Firstly, an 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS) working mother solution with equal volumes of ABTS solution and oxidant solution was prepared. Then the radical cation (ABTS+•) solution was prepared (7 mmol/L ABTS in water) with potassium persulfate (2.45 mmol/L) and incubated at room temperature for 12~16 h, to achieve a stable oxidative state in a space without light. The ABTS+• solution was diluted with phosphate-buffered saline (PBS) (pH 7.4) to obtain a solution with an absorbance of (0.700±0.01) at 734 nm. A portion of 10 μ L of sample mixed with an aliquot of 200 μ L of ABTS solution reacted for 2~6 min in the dark space at room temperature. And the accumulations of oxidation products were measured by the absorption spectrum of 734 nm with a UV-spectrophotometer. The absorbance of ABTS+• blank and trolox as a positive control was also measured at the paralleled approach. Percentage inhibitions of ABTS were calculated according to the following equation: Inhibition of *A*₇₃₄(%)=(1−*A*_s/*A*_b)×100%, where *A*_s=absorbance of sample, *A*_b=absorbance of blank.

Supporting Information 1D NMR and 2D NMR, HR-ESI-MS spectra of compound **1**. The Supporting Information is available free of charge via the Internet at <http://sioc-journal.cn>.

References

- [1] Huang, H. C.; Lin, M. K.; Hwang, S. Y.; Hwang, T. L.; Kuo, Y. H.; Chang, C. I.; Ou, C. Y.; Kuo, Y. H. *Molecules* **2013**, *18*, 8752.
- [2] Zhao, T.; Nong, X. H.; Zhang, B.; Tang, M. M.; Huang, D. Y.; Wang, J. L.; Xiao, J. L.; Chen, G. Y. *Phytochem. Lett.* **2020**, *36*, 115.
- [3] Yu, X. H.; Li, P. P.; Geng, Y. Y.; Huang, Y. J.; Hua, Y. *Chem. Ind. Forest Prod.* **2017**, *37*, 149 (in Chinese).
(余小红, 李盼盼, 耿圆圆, 黄艳杰, 华燕, 林产化学与工业, **2017**, *37*, 149.)
- [4] Tan, S. B.; Guo, T.; Tang, X. F.; Song, T. T.; Wang, Y. *Chem. Nat. Compd.* **2018**, *54*, 869.
- [5] Zhao, C. C.; Shao, J. H.; Li, X. *China J. Chin. Mater. Med.* **2009**, *34*, 2197 (in Chinese).
(赵春超, 邵建华, 李锐, 中国中药杂志, **2009**, *34*, 2197.)
- [6] Sun, Y. H.; Ling, Y.; Ren, M. R.; Zhou, X. G.; Wang, J. Y.; Ma, Y. L. *Chin. Tradit. Herbal Drugs* **2010**, *41*, 1418 (in Chinese).
(孙勇慧, 凌勇, 任美荣, 周学刚, 王佳瑜, 马英丽, 中草药, **2010**, *41*, 1418.)
- [7] Ou, Y. W.; Luo, Y. F.; Cheng, S. J.; Zhang, Y. K.; Tang, C. Y.; Tang, D. F.; Tang, M.; Xiao, S.; He, Y. T.; Xiong, G. H.; Li, S. X. *Chin. Tradit. Herbal Drugs* **2018**, *49*, 3208 (in Chinese).
(欧阳文, 罗懿钊, 程思佳, 张云坤, 唐纯玉, 唐代风, 唐敏, 肖珊, 贺玉婷, 熊广华, 李顺祥, 中草药, **2018**, *49*, 3208.)
- [8] Cheng, Z. Y.; Zhou, L.; Liu, S. F.; Bai, M.; Song, S. J.; Huang, X. X. *J. Shenyang Pharm. Univ.* **2015**, *32*, 844 (in Chinese).
(程卓阳, 周乐, 刘思帆, 白明, 宋少江, 黄肖霄, 沈阳药科大学学报, **2015**, *32*, 844.)

- [9] Sun, K.; Li, X.; Li, W.; Wang, J. H.; Liu, J. M.; Sha, Y. *Chem. Pharm. Bull.* **2004**, *52*, 1483.
- [10] Nicolás, G.; María, M. Z.; Ariel, M. S. *J. Org. Chem.* **2015**, *80*, 12526.
- [11] Mustafa, O.; Reese, R. N.; Artemio, Z. T. J.; Joseph, C. S.; Miller, A. R. *J. Agric. Food Chem.* **2006**, *54*, 1151.
- [12] Cai, J.; Nong, X. H.; Wang, B.; Zeng, W. N.; Huang, G. L.; Zhang, Z. Y.; Zheng, C. J. *Chin. J. Org. Chem.* **2021**, *41*, 2905 (in Chinese). (蔡瑾, 农旭华, 王斌, 曾尾女, 黄国雷, 张子怡, 郑彩娟, 有机化学, **2021**, *41*, 2905.)

(Lu, Y.)